



<http://dx.doi.org/10.11646/zootaxa.3866.2.6>

<http://zoobank.org/urn:lsid:zoobank.org:pub:F66AC418-C598-443C-9774-CE1C0F25E2F6>

Pratylenchus quasitereoides n. sp. from cereals in Western Australia

MIKE HODDA^{1,2}, SARAH J. COLLINS³, VIVIEN A. VANSTONE³, DIANA HARTLEY¹,
WOLFGANG WANJURA⁴ & MONICA KEHOE³

¹Australian National Insect Collection, CSIRO National Research Collections Australia, GPO Box 1700, Canberra, ACT, 2601 Australia. E-mail: mike.hodda@csiro.au, diana.hartley@csiro.au

²CSIRO Biosecurity Flagship, EcoSciences Precinct, GPO Box 2583, Brisbane QLD 4001 Australia

³Department of Agriculture and Food Western Australia, LMB 4, Bentley Delivery Centre, WA, 6983 Australia.

E-mail: sarah.collins@agric.gov.au, vavanstone@bigpond.com, monica.kehoe@agric.gov.au

⁴Atlas of Living Australia, CSIRO National Research Collections Australia, GPO Box 1700, Canberra, ACT, 2601.

E-mail: wolf.wanjura@csiro.au

Abstract

Pratylenchus quasitereoides n. sp. is described from Western Australia. It is characterized by 2 external incisures in the head cuticle, 4 lateral incisures at mid body, stylet length 17 µm to 19 µm, *V* greater than 75%, PUS less than 2 body diameters long and crenate tail terminus. Molecular data confirm the separation of the new species from morphologically similar and sympatric congeners. The host range also differs from *P. teres* as well as the sympatric *P. neglectus*, *P. thornei* and *P. penetrans*. Reproduction rates on oat and lupin differed between the new species and *P. neglectus*. The species was originally described as *P. teres*, but the species concept of *P. teres* now encompasses a considerable range of different attributes spread over two described subspecies and three variant populations. The new species differs from all these subspecies and populations in at least two characters. It differs from all populations of *P. teres* most notably in having four rather than 6 lateral lines and a more posterior vulva. It differs from *P. teres vandebergae* in having a longer stylet and longer overlap of the intestine by the oesophageal glands. Characters which can be used under low magnification to separate the new species from the closest sympatric congeners (*P. thornei* and *P. crenatus*) are discussed.

Key words: 28S, *Avena sativa*, cereal, D3, host, lupin, *Lupinus angustifolius*, molecular biology, morphology, oat, *P. teres*, *P. neglectus*, *P. thornei*, reproduction rate, Root Lesion Nematode, rRNA, taxonomy

Introduction

Recently, *Pratylenchus teres* Khan and Singh 1974 was redescribed from Western Australia (Riley & Wouts 2001), having been originally described from Punjab, India. It was also redescribed from the Caribbean and South Africa at about the same time as Western Australia (Carta *et al.* 2002; van den Berg & Queneherve, 2000). Since the original find in Western Australia, many additional specimens have been located which allow a better evaluation of the variability in characteristics among the Western Australian populations relative to the variability in the *P. teres* found elsewhere. They have also given better understanding of the differences between the Western Australian populations and the *P. teres* found elsewhere. These differences are sufficient to differentiate the Western Australian population as a new species, which is described herein as *Pratylenchus quasitereoides* n.sp. Phylogenetic analyses of 28S rRNA also suggest the new species is separate from *P. teres* and other morphologically-similar congeners, as well as sympatric congeners. There are also differences between the new species and others in host relationships.

Material & methods

Specimens of the new species were obtained from several collections between 1998 and 1999, including the first

Multiplication factors for *P. neglectus* and *P. thornei* on lupin and oats are generally less than 1, but generally greater than 1 for *P. quasitereoides* n. sp. (Vanstone *et al.* 2005). On the same varieties of lupin and oats, reproduction rates of *P. quasitereoides* n. sp. (1.6 and 8.1, respectively) were much greater than those of *P. neglectus* (1.0 and 1.6, respectively). However, different varieties have different host status to *P. quasitereoides* n.sp. as well as for *P. neglectus* and *P. thornei* (Collins *et al.* 2013, Vanstone 2007, Vanstone *et al.* 2005).

Etymology. The name indicates the similarity of the new species with *P. teres* from the suffix “oides”, and that the species has been known as *P. teres* “quasi”. The species epithet is mostly Latin neuter, but the suffix is Greek (as with the generic name), and chosen for euphony above the Latin “similis”.

TABLE 3. Hosts of the most common species of *Pratylenchus* in Western Australia, plus *P. teres* sensu stricto.

Host	<i>P. quasitereoides</i> n. sp.	<i>P. teres</i>	<i>P. neglectus</i>	<i>P. thornei</i>	<i>P. penetrans</i>
barley (<i>Hordeum vulgare</i> L.)	X		X	X	X
wheat (<i>Triticum aestivum</i> L.)	X		X	X	X
oat (<i>Avena sativa</i> L.)	X		X		X
chickpea (<i>Cicer arietinum</i> L.)	X		X	X	X
lupin (<i>Lupinus angustifolius</i> L.)	X				X
canola (<i>Brassica napus</i> L.)	X		X	X	X
potato (<i>Solanum tuberosum</i> L.)		X			
mustard (<i>Brassica juncea</i> L.)		X	X	X	
safflower (<i>Carthamus tinctorius</i> L.)		X			
cotton (<i>Gossypium hirsutum</i> L.)		X			
pearl millet (<i>Pennisetum glaucum</i> (L.))		X			
sugar cane (<i>Saccharum officinarum</i> L.)		X			
tobacco (<i>Nicotiana tabacum</i> L.)		X			
medic (<i>Medicago</i> spp.)			X		
durum (<i>Triticum durum</i> L.)			X		X
common vetch (<i>Vicia sativa</i> L.)			X	X	
field pea (<i>Pisum sativum</i> L.)					X
fabo bean (<i>Vicia faba</i> l.)					X
triticale (<i>Triticum x Secale</i>)					X

References

- Braun, A.L. & Loof, P.A.A. (1966) *Pratylenchoides laticuada* n. sp., a new endoparasitic phytonematode. *Netherlands Journal of Plant Pathology*, 72, 241–245.
<http://dx.doi.org/10.1007/BF02650210>
- Cadet, P., van den Berg, E., Delatte, A. & Fiard, J.-P. (1994) Comparaison de quelques peuplements nematologiques des Petites Antilles. *Biogeographica*, 70, 125–138.
- Carta, L.K., Handoo, Z.A., Skantar, A.M., van Biljon, J. & Botha, M. (2002) Redescription of *Pratylenchus teres* Khan & Singh, 1974 (Nemata: Pratylenchidae), with the description of a new subspecies from South Africa, and a phylogenetic analysis of related species. *African Plant Protection*, 8, 13–24.
- Cobb, N.A. (1917) A new parasitic nema found infesting cotton and potatoes. *Journal of Agricultural Research*, 11, 27–33.
- Collins, S.J., Kelly, S., Hunter, H., MacLeod, B., Debrincat, L., Teasdale, J., Versteeg, C. & Zhang, Z. (2013) *P. teres*—WA’s home grown Root Lesion Nematode (RLN) and its unique impacts on broadacre crops. 2013 WA Crop Updates, GRDC & DAFWA, Perth., 4 pp.
- Department of Agriculture Western Australia (2005) *Annual Report 2003–2004*. Western Australian Department of Agriculture, Perth, 121 pp.
- Felsenstein, J. (1985) Confidence-limits on phylogenies—an approach using the bootstrap. *Evolution*, 39, 783–791.
<http://dx.doi.org/10.2307/2408678>

- Filipjev, I.N. & Schuurmans Stekhoven, J.H. (1941) *A manual of agricultural helminthology*. E. J. Brill, Leiden, 878 pp.
- Geraert, E. (1968) Morphometric relations in nematodes. *Nematologica*, 14, 171–182.
<http://dx.doi.org/10.1163/187529268X00390>
- Hodda, M. & Nobbs, J.M. (2008) A review of current knowledge on particular taxonomic features of the Australasian nematode fauna, with special emphasis on plant feeders. *Australasian Plant Pathology*, 37, 308–317.
<http://dx.doi.org/10.1071/AP08024>
- Hooper, D.J. (1986a) Extraction of free-living stages from soil. In: Southey, J.F. (Ed.), *Laboratory methods for work with plant and soil nematodes*. 6th ed. Her Majesty's Stationery Office, London, pp. 5–30.
- Hooper, D.J. (1986b) Drawing and measuring nematodes. In: Southey, J.F. (Ed.), *Laboratory methods for work with plant and soil nematodes*. 6th ed. Her Majesty's Stationery Office, London, pp. 87–94.
- Karssen, G. & Bolk, R.J. (2000) An additional character useful for the identification of *Pratylenchus crenatus* Loof 1960 (Nematoda: Pratylenchidae). *Nematology*, 2, 695–697.
<http://dx.doi.org/10.1163/156854100509556>
- Khan, E. & Singh, D.B. (1974) Five new species of *Pratylenchus* (Nematoda : Pratylenchidae) from India. *Indian Journal of Nematology*, 4, 199–211.
- Loof, P.A.A. (1992) The family Pratylenchidae Thorne, 1949. In: Nickle, W.R. (Ed.), *Manual of Agricultural Nematology*. Marcel Dekker, New York, pp 363–421.
- Nei, M. & Kumar, S. (2000) *Molecular Evolution and Phylogenetics*. Oxford University Press, New York, 333 pp.
- Nunn, G.B., Theisen, B.F., Christensen, B. & Arctander, P. (1996) Simplicity-correlated size growth of the nuclear 28S ribosomal RNA D3 expansion segment in the crustacean order Isopoda. *Journal of Molecular Evolution*, 42, 211–223.
<http://dx.doi.org/10.1007/BF02198847>
- Palomares-Rius, J.E., Castillo, P., Liebana, G., Vovlas, N., Landa, B.B., Navas-Cortes, J.A. & Subbotin, S.A. (2010) Description of *Pratylenchus hispaniensis* n. sp from Spain and considerations on the phylogenetic relationship among selected genera in the family Pratylenchidae. *Nematology*, 12, 429–451.
<http://dx.doi.org/10.1163/138855409X12559479585043>
- Rensch, B. (1924) *Aphelenchus neglectus* sp. n., eine neue parasitare Nematodenart. *Zoologischer Anzeiger Leipzig*, 59, 277–280.
- Riley, I.T. & Kelly, S.J. (2002) Endoparasitic nematodes in cropping soils of Western Australia. *Australian Journal of Experimental Agriculture*, 42, 49–56.
<http://dx.doi.org/10.1071/EA01054>
- Riley, I.T. & Wouts, W.M. (2001) *Pratylenchus* and *Radopholus* species in agricultural soils and native vegetation in Southern Australia. *Transactions of the Royal Society of South Australia*, 125, 147–153.
- Ryss, A.Y. (1982) [New phyto nematode species of the genus *Pratylenchus* in the Estonian SSR, USSR.] *Eesti NSV Teaduste Akadeemia Toimetised Bioloogia*, 31, 22–29.
- Ryss, A.Y. (2002) Genus *Pratylenchus* Filipjev: multientry and monoentry keys and diagnostic relationships (Nematoda: Tylenchida: Pratylenchidae). *Zoosystematica Rossica*, 10, 241–255.
- Rzhetsky, A. & Nei M. (1992) A simple method for estimating and testing minimum-evolution trees. *Molecular Biology and Evolution*, 9, 945–967.
- Saitou, N. & Nei, M. (1987) The neighbor-joining method: A new method for reconstructing phylogenetic trees. *Molecular Biology and Evolution*, 4, 406–425.
- Seinhorst, J. W. (1959) A rapid method for the transfer of nematodes from fixative to anhydrous glycerine. *Nematologica*, 4, 67–69.
<http://dx.doi.org/10.1163/187529259X00381>
- Sher, S.A., Allen M.W. (1953) Revision of the genus *Pratylenchus* (Nematoda: Tylenchidae). *University of California Publications in Zoology*, 57, 441–470.
- Siddiqi, M.R. (2000) *Tylenchida: Parasites of plants and insects*. CABI, New York, 833 pp.
- Subbotin, S.A., Ragsdale, E.J., Mullens, T., Roberts, P.A., Mundo-Ocampo, M. & Baldwin, J.G. (2008) A phylogenetic framework for root lesion nematodes of the genus *Pratylenchus* (Nematoda): Evidence from 18S and D2-D3 expansion segments of 28S ribosomal RNA genes and morphological characters. *Molecular Phylogenetics and Evolution*, 48, 491–505.
<http://dx.doi.org/10.1016/j.ympev.2008.04.028>
- Taheri, Z.M., Maafi, Z.T., Subbotin, S.A., Pourjam, E. & Eskandari, A. (2013) Molecular and phylogenetic studies on Pratylenchidae from Iran with additional data on *Pratylenchus delattrei*, *Pratylenchoides alkani* and two unknown species of *Hirschmanniella* and *Pratylenchus*. *Nematology*, 15, 633–651.
- Tamura, K., Nei M. & Kumar, S. (2004) Prospects for inferring very large phylogenies by using the neighbor-joining method. *Proceedings of the National Academy of Sciences (USA)*, 101, 11030–11035.
<http://dx.doi.org/10.1073/pnas.0404206101>
- Tamura, K., Peterson, D., Peterson, N., Stecher, G., Nei, M. & Kumar, S. (2011) MEGA5: Molecular Evolutionary Genetics Analysis using Maximum Likelihood, Evolutionary Distance, and Maximum Parsimony Methods. *Molecular Biology and Evolution*, 28, 2731–2739.
<http://dx.doi.org/10.1093/molbev/msr121>

- Tamura, K. & Nei, M. (1993) Estimation of the number of nucleotide substitutions in the control region of mitochondrial DNA in humans and chimpanzees. *Molecular Biology and Evolution*, 10, 512–526.
- Thompson, J.D., Higgins, D.G. & Gibson, T.J. (1994). CLUSTAL W: improving the sensitivity of progressive multiple sequence alignment through sequence weighting, position-specific gap penalties and weight matrix choice. *Nucleic Acids Research*, 22, 4673–4680.
<http://dx.doi.org/10.1093/nar/22.22.4673>
- Tobar-Jimenez, A. (1963) *Pratylenchoides guevarai* sp. nuevo, nematode tylenchido relacionado con el cipres (*Cupressus sempervirens* L.). *Revista Iberica de Parasitologia*, 23, 27–36.
- van den Berg, E. & Queneherve, P. (2000) *Hirschmanniella caribbeana* sp. n. and new records of *Pratylenchus* spp. (Pratylenchidae: Nematoda) from Guadeloupe, French West Indies. *Nematology*, 2, 179–190.
<http://dx.doi.org/10.1163/156854100509079>
- Vanstone, V. A. (2007) Root Lesion and Burrowing Nematodes in Western Australian cropping systems. *DAFWA Bulletin*, 4698, 1–19.
- Vanstone, V.A., Kelly, S.J., Hunter, H.F. & Gilchrist, M.C. (2005) *Rotations for nematode management*. 2005 WA Crop Updates, GRDC & DAFWA, Perth, 7 pp.
- Whitehead, A.G. & Hemming, J.R. (1965) A comparison of some quantitative methods of extracting small vermiform nematodes from soil. *Annals of Applied Biology*, 55, 25–38.
<http://dx.doi.org/10.1111/j.1744-7348.1965.tb07864.x>