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A new species of *Tozzita* Kramer (Hemiptera: Cicadellidae: Neocoelidiinae) and first record of *Tozzita ips* Kramer from Brazil

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Abstract

A new species of *Tozzita* Kramer, 1964, *Tozzita bifida* sp. nov., is described and illustrated. The new species can be distinguished from the other species of the genus by the following characteristics: (1) forewing with commissural brown band; (2) pygofer with dentiform process on ventral margin and apex not spatulate; (3) subgenital plates with macrosetae at apex; (4) style with preapical lobe developed and produced posteriorly; (5) aedeagus symmetrical; (6) anal tube with bifid basal process. *Tozzita ips* Kramer, 1964 is firstly recorded from Brazil. Additionally, an updated genus description and an identification key to males of the species are provided.

Key words: *Tozzita bifida* sp. nov., leafhoppers, identification key, Neotropical region

Introduction

The New World leafhopper subfamily Neocoelidiinae comprises 174 valid species in 33 genera (Gonçalves *et. al.* 2011, 2012a, b; Marques-Costa & Cavichioli 2012). The subfamily is mainly Neotropical, with only four genera recorded from the Nearctic Region: *Coelella* DeLong, 1953, *Neocoelidia* Gillette & Baker, 1895, *Cocoelidia* DeLong, 1953, and *Neocoelidiana* DeLong, 1953 (Nielson & Knight 2000).

Tozzita was described by Kramer (1964) based on the type-species, *T. ips* Kramer, 1964, from Bolivia. Later in 1967, Kramer described *T. petulans* from Brazil. Herein, a new species *T. bifida* sp. nov. is described from the type-locality Ipixuna, Amazonas, Brazil. In the same locality, one male of *T. ips* was also collected, representing the first record of this species in Brazil.

In this paper we propose modifications to the generic description of *Tozzita* made ??by broadening the diagnostic characters of the genus for inclusion of *Tozzita bifida* sp. nov. Additionally, an identification key to males of the species are provided.

Material and methods

For observation of genital structures, the abdomen was removed and placed in hot 10% KOH, following Oman (1949), with small modifications in heating time (decreased to about 3-5 minutes). Softened genitalia were washed for 5–10 minutes in hot water and placed on a depression slide with glycerin to take photographs. After being photographed, the genitalia were kept in plastic microvials with glycerin, and pinned together with the specimen. Photographs were taken with a camera attached to a Leica stereo microscope, using the stacking image-capturing computer software LAS Montage.