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# Neocordana gen. nov., the causal organism of Cordana leaf spot on banana

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### Abstract

Cordana leaf spot of banana is shown to be associated with several species of a new genus described here as *Neocordana gen. nov.* Furthermore, *Neocordana* belongs to *Pyriculariaceae* (*Magnaporthales*) rather than *Cordanaceae* where the type species of *Cordana*, *C. pauciseptata* resides. *Neocordana* is established to accommodate *Cordana musae*, *C. johnstonii*, *C. versicolor*, and a previously undescribed species, *N. musicola*, which is morphologically and phylogenetically distinct. *Neocordana* species are found to be associated with leaves of *Musa* spp. (*Musaceae*) and *Canna denudata* (*Cannaceae*). Based on these results, *Cordanaceae* is best recognized in a separate order, established here as *Cordanales ord. nov*.

Key Words: Cordanales, Magnaporthales, Musa, plant pathogenic fungi, Pyriculariaceae, systematics

## Introduction

Cordana leaf spot is a common and widespread disease on banana and plantain. Although it is considered as a minor pathogen of banana, it can cause serious defoliation of plantains in Central America during and following periods of wet weather (Jones 1999, Ploetz et al. 2003). Cordana leaf spot is characterised by large, pale brown, oval to fusiform necrotic lesions with pale grey concentric rings, with a dark brown border surrounded by a bright yellow halo, separating lesions from healthy leaf tissue (Jones 1999). Most damage occurs when the pathogen gains entry to leaf tissue weakened because of age, adverse environmental conditions, nutritional deficiencies, wounds or through lesions caused by other pathogens. The leaves ultimately turn brown and dry out (Jones 1999). This is especially apparent when the disease occurs together with Sigatoka leaf spots (Arzanlou et al. 2008), in which case the lesions may encompass the entire leaf margin and large portions of the lamina can be affected (Ploetz et al. 2003). Leaf infection normally occurs at night during rainy periods or when dew is present. The conidia germinate in a film of moisture on the leaf surface and after a few hours appressoria are formed. The appressoria enable the fungus to penetrate into the host epidermal cells. Species most commonly associated with Cordana leaf spot of banana include Cordana musae (Zimm.) Höhn. (1923: 60) and C. johnstonii M.B. Ellis (1971: 125). Cordana leaf spot has been reported mainly on M. sapientum L. but also can be found on M. acuminata ssp. banksii N.W.Simmonds, M. acuminata ssp. banksii × M. schizocarpa N.W.Simmonds, M. balbisiana Colla, M. boman Argent, M. maclayi F.Muell, M. schizocarpa N.W.Simmonds and Enset glaucum (Roxb.) Cheesman (Jones, 1999). Another plant pathogenic species, Cordana versicolor D.J. Soares & R.W. Barreto (2005: 18), was described from Brazil causing eye-spot disease on leaves of Canna denudata (Soares et al. 2005).

The asexual genus *Cordana* was erected by Preuss (1851) with *C. pauciseptata* Preuss (1851: 129) as type species. Species of *Cordana* are characterized by brown, septate conidiophores with swollen conidiogenous zones, terminal and intercalary conidiogenous cells, and pale brown to brown, 0–1-septate conidia. Ecologically, *Cordana* species can be separated into two groups, namely phanerogam leaf-inhabiting species (saprobic or pathogenic) or saprobic on rotten

clade in the family *Cordanaceae*, as the sister clade of *Coniochaetales* in *Sordariomycetes*. *Cordanales* is here erected for the single family *Cordanaceae*. On the other hand, *Neocordana* is introduced to accommodate phytopathogenic species similar to *Cordana* but closely related to *Pyriculariaceae* (*Magnaporthales*) rather than *Cordanaceae*. The family *Pyriculariaceae* was recently introduced by Klaubauf *et al.* (2014) and includes important plant pathogens along with *Deightoniella* S. Hughes (1952: 48) and several pyricularia-like genera. *Deightoniella* can be easily differentiated from *Neocordana* since it produces conidiophores reduced to conidiogenous cells with a flattened scar, and conidia with a central pore in the base (Hughes 1952, Klaubauf *et al.* 2014), while *Neocordana* has septate conidiophores with denticulate conidiogenous cells and conidia with a protruding hilum. On the other hand, pyricularia-like fungi and *Neocordana* are similar in having brown, septate conidiophores with polyblastic, denticulate conidiogenous cells. Nevertheless, pyricularia-like fungi are different from *Neocordana* in having pyriform to obclavate, 2-septate conidia, and grow on grasses and other plants (Seifert *et al.* 2011, Klaubauf *et al.* 2014). *Neocordana* differs by having broadly ellipsoid, obovoid to pyriform, 1-septate conidia and are pathogenic to species of *Musa* or *Canna*.

In the phylogenetic analysis generated here (Figure 1), *Neocordana* is represented by *N. musae* and *N. musicola*. *N. musae* is designated as the type species of the genus, and *N. musicola* is introduced as a new species, similar but different from *N. musae*, based on morphological and molecular data. Unfortunately cultures of *N. johnstonii* and *N. versicolor* were not available to include in the phylogenetic analysis. Nevertheless we propose new combinations for *N. johnstonii*, and *N. versicolor*, based on morphology, pathogenicity and host association. Further studies are needed to confirm their phylogenetic affinities in *Neocordana*.

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#### References

Arzanlou, M., Groenewald, J.Z., Fullerton, R.A., Abeln, E.C.A, Carlier, J., Zapater, M.-F., Buddenhagen, I.W., Viljoen, A. & Crous, P.W. (2008) Multiple gene genealogies and phenotypic characters differentiate several novel species of *Mycosphaerella* and related anamorphs on banana. *Persoonia* 20: 19–37.

http://dx.doi.org/10.3767/003158508X302212

- Crous, P.W., Gams, W., Stalpers, J.A., Robert, V. & Stegehuis, G. (2004) MycoBank: an online initiative to launch mycology into the 21st century. *Studies in Mycology* 50:19–22.
- Crous P.W., Schoch C.L., Hyde K.D., Wood, A.R., Gueidan, C., de Hoog, G.S. & Groenewald, J.Z. (2009a) Phylogenetic lineages in the Capnodiales. *Studies in Mycology* 64: 17–47. http://dx.doi.org/10.3114/sim.2009.64.02
- Crous, P.W., Verkley, G.J.M., Groenewald, J.Z. & Samson, R.A. (Eds) (2009b) *Fungal Biodiversity*. [CBS Laboratory Manual Series no. 1.] Utrecht: CBS-KNAW Fungal Biodiversity Centre.
- Ellis, M.B. (1971) Dematiaceous hyphomycetes. X. Mycological Papers 125: 1-30.
- de Hoog, G.S. (1973) A new species of Cordana (Dematiaceae, Hyphomycetes). Acta Botanica Neerlandica 22: 209-212.
- de Hoog, G.S., van, Oorschot, C.A.N. & Hijwegen, T. (1983) Taxonomy of the *Dactylaria* complex. II. *Dissoconium gen. nov.* and *Cordana* Preuss. *Proceedings of the Koninklijke Nederlandse Akademie van Wetenschappen, Series C* 86: 197–206.
- Hernández-Restrepo, M., Gené, J., Mena-Portales, J., Cano, J., Madrid, H., Castañeda-Ruiz, R.F. & Guarro, J. (2014) New species of *Cordana* and epitypification of the genus. *Mycologia* 106: 723–734. http://dx.doi.org/10.3852/13-122
- Hughes, S.J. (1952) Fungi from the Gold Coast. I. Mycological Papers 48: 1-91.
- Huhndorf, S.M., Miller, A.N. & Fernández, F.A. (2004) Molecular systematics of the Sordariales: the order and the family Lasiosphaeriaceae redefined. *Mycologia* 96: 368–387.
  - http://dx.doi.org/10.2307/3762068
- Katoh, K. & Standley, D.M. (2013) MAFFT multiple sequence alignment software version 7: improvements in performance and usability. *Molecular Biology and Evolution* 30: 772–80. http://dx.doi.org/10.1093/molbev/mst010

- Jones, D.R. (1999) Fungal diseases of the foliage. Cordana leaf spot. *In:* Jones, D.R. (Ed.) *Diseases of Banana, Abaca and Enset.* CABI Publishing. Wallingford, UK, pp 99–101.
- Nylander, J.A.A. (2004) MrModeltest v2.2. Uppsala: distributed by the author. Evolutionary Biology Centre, Uppsala University.

Ploetz, R.C., Thomas, J.E. & Slabaugh, W.R. (2003) Diseases of Banana and Plantain. In: Ploetz, R.C. (Ed.) Diseases of Tropical Fruit Crops. CABI Publishing. Wallingford, UK, 543 pp.

http://dx.doi.org/10.1079/9780851993904.0073

Preuss, C.G.T. (1851) Übersicht untersuchter Pilze, besonders aus der Umgegend von Hoyerswerda. Linnaea 24: 99–153.

Priest, M.J. (1990) Distribution of Cordana species on Musa in Australia. Mycological Research 94: 861-863.

http://dx.doi.org/10.1016/S0953-7562(09)81394-4

Rayner, R.W. (1970) A mycological colour chart. Kew Commonwealth Mycological Institute.

- Réblová, M., Barr, M.E. & Samuels, G.J. (1999) *Chaetosphaeriaceae*, a new family for *Chaetosphaeria* and its relatives. *Sydowia* 51: 49–70.
- Réblová, M. & Seifert, K.A. (2007) A new fungal genus, *Teracosphaeria*, with a phialophora-like anamorph (Sordariomycetes, Ascomycota). *Mycological Research* 3: 287–298.

http://dx.doi.org/10.1016/j.mycres.2006.12.005

Réblová, M. & Winka, K. (2000) Phylogeny of *Chaetosphaeria* and its anamorphs based on morphological and molecular data. *Mycologia* 92: 939–954.

http://dx.doi.org/10.2307/3761589

Ronquist, F., Teslenko, M., Mark, P. van der, Ayres, D.L., Darling, A., Höhna, S., Larget, B., Liu, L., Suchard, M.A. & Huelsenbeck, J.P. (2012) MrBayes 3.2: efficient Bayesian phylogenetic inference and model choice across a large model space. *Systematic Biology* 61: 539–542.

http://dx.doi.org/10.1093/sysbio/sys029

- Seifert, K., Morgan-Jones, G., Gams, W. & Kendrick, B. (2011). *The Genera of Hyphomycetes*. [CBS Biodiversity Series 9]. Utrecht: CBS-KNAW Fungal Biodiversity Centre.
- Soares, D.J., Nechet, K.L. & Barreto, R.W. (2005) Cordana versicolor sp. nov. (dematiaceous hyphomycete) causing leaf-spot on Canna denudata (Cannaceae) in Brazil with observations on Cordana musae. Fungal Diversity 18: 147–155.
- Stamatakis, A., Hoover, P. & Rougemont, J. (2008) A rapid bootstrap algorithm for the RAxML web-servers. *Systematic Biology* 75: 758–771.

http://dx.doi.org/10.1080/10635150802429642

Tamura, K., Stecher, G., Peterson, D., Filipski, A. & Kumar, S. (2013) MEGA6: Molecular evolutionary genetics analysis version 6.0. Molecular Biology and Evolution 30: 2725–2729.

http://dx.doi.org/10.1093/molbev/mst197

- Vilgalys, R. & Hester, M. (1990) Rapid generic identification and mapping enzymatically amplified ribosomal DNA from several *Cryptococcus* species. *Journal of Bacteriology* 172: 4238–4246.
- von Höhnel, F. (1923) Studien über hyphomyzeten. Centralblatt für Bakteriologie, Parasitenkunde und Infektionskrankheiten Abteilung. 60: 1–23.
- White, T.J., Bruns, T., Lee, J. & Taylor, S.B. (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. *In:* Innis, M.A., Gelfand, D.H., Sninsky, J.J. & White, T.J. (Eds.) *PCR Protocols: A Guide to Methods and Applications*. Academic Press, San Diego, pp. 315–322.

http://dx.doi.org/10.1016/B978-0-12-372180-8.50042-1

Zelski, S.E., Balto, J.A., Do, C., Raja, H.A., Miller, A.N. & Shearer, C.A. (2014) Some dematiaceous freshwater microfungi from Peru and their previously unknown phylogenetic relationships based on 28S nrDNA, and reports of related taxa based on morphology. *IMA Fungus* 5: 425–438.

http://dx.doi.org/10.5598/imafungus.2014.05.02.07

Zimmermann, A. (1902) Ueber einige an tropischen kulturepflanzen beobachtete pilze. II. *Centralblatt fur Bakteriologie, Parasitenkunde und Infektionskrankheiten II Abteilung.* 8: 216–221.